# Deuterium Depletion can Decrease the Expression of C-myc Ha-ras and p53 Gene in Carcinogen-Treated Mice

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Abstract. In spite of the fact that the deuterium concentration is over 10 mmol/l in all living organisms, its possible role has been ignored for six decades. Recent studies have shown that the depletion of the naturally occurring deuterium can result in tumour regression in mice, dogs, cats and humans. The effect of deuterium depletion on gene expression plays a key part in tumour development. The carcinogen, 7,12-dimethylbenz(a)anthracene (DMBA), was used to increase gene expression in "short term" investigations. The expression of c-myc, Ha-ras and p53 gene was followed in CBA/Ca sensitive inbred mice drinking tap water or deuterium-depleted water (DDW) after induction. By detecting the RNA expression 48 hours after exposure to the carcinogen it was found that the expression of all genes investigated was inhibited in six different organs (spleen, lung, thymus, kidney, liver and lymph node) in the DDW-treated group. It is suggested that genes playing a key role in the cell cycle regulation and tumour development are sensitive to deuterium depletion.

It has been known for decades that the mass difference between hydrogen and deuterium leads to differences in the physical and chemical behaviour of the two stable isotopes (1-2). The effect of the replacement of hydrogen with deuterium (e.g. by the application of heavy water, D<sub>2</sub>O) is also well documented (3-5). The possible role of naturally occurring deuterium, whose concentration is over 16 mmoi/l in surface water and 12-14 mmol/l in living organisms (6), in biological systems was first investigated in the early nineties. The results showed that the partial replacement of the naturally occurring deuterium with hydrogen by the application of deuterium-depleted water (DDW) inhibited cell growth rate (L<sub>929</sub> fibroblast cell line) and caused complete tumour regression in xenotransplanted mice (7). Subsequent experiments revealed

Abbreviations: DMBA, 7,12-dimethylbenz(a)anthracene, DDW, deuterium-depleted water.

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Key Words: Deuterium, deuterium depletion, deuterium-depleted water, DMBA, c-myc, Ha-ras, p53, gene expression.

that deuterium depletion inhibited the proliferation of tumorous cell lines *in vitro* (PC-3 human prostate, MDA-MB-231, MCF-7 human breast and M14 human melanoma), caused regression of breast adenocarcinoma in dogs and showed efficacy in Phase II double blind clinical trial with human prostate cancer (8). The

application of DDW also triggered apoptosis in vitro (8) and in vivo (9).

The *in vivo* detection of the expression of certain oncogenes and tumour suppressor genes in the "short term" experiment is a good model for the early detection of exposure to carcinogens (10-28). In order to investigate the possible influence of deuterium depletion on the expression of genes involved in malignant transformation, 7,12-dimethyl benz( $\alpha$ )anthracene (DMBA) was used to increase the expression of (proto)oncogenes and tumour suppressor gene in sensitive (10) inbred mice.

The present study was designed to examine whether deuterium depletion influence the expression of the c-myc, Ha-ras and p53 genes.

## **Materials and Methods**

Animals. 6 to 8-week-old CBA/Ca (Lati, Gödölló, Hungary) inbred mice were kept in a conventional breeding house.

Treatment. Each group was treated with a single dose of 40 mg/bwkg 7,12-dimethylbenz( $\alpha$ )anthracene, which was dissolved in corn oil and given intraperitoneally. The drinking water of the animals in the treated group (24 mice) was replaced for two days with DDW (21 ppm  $\pm$  3 ppm deuterium) after the injection of DMBA. The animals (12 mice) in the control group consumed tap-water (150 ppm  $\pm$  3 ppm).

Gene expression. 48 hours after the gene induction with DMBA and the subsequent treatment with DDW the animals were autopsied. For RNA isolation the lung, liver, kidney, thymus, spleen and lymph nodes were removed and pooled into two-two groups. The RNA was isolated with Trisol (GIBCO, Grand Island, NY, USA) according to manufacturer's instructions (29). RNA samples after OD control (A260/A280>1.8) were diluted to 0.1 μg/μl and blotted 100 μl onto Hybond N+ membrane (Amersham, Little Chalfont, UK) so that each blot contained 10 μg of total RNA. After heat fixation the membranes were hybridized with chemiluminescently labeled cloned DNA probes of c-myc, Ha-ras and p53 (American Tissue Culture Collection Manassas, USA). The hybridization protocol was carried out according to the manufacturers' instructions (ECL gene detection system, Amersham) (30). The signals

#### Gene expression

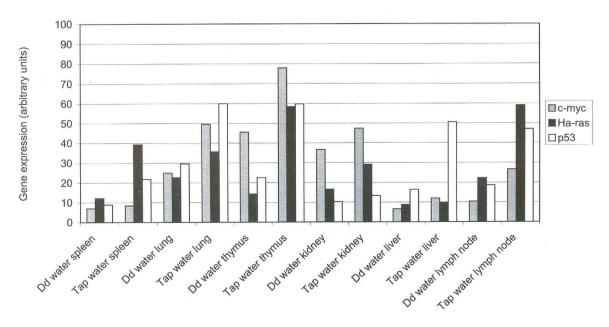


Figure 1. Deuterium depletion inhibited, DMBA induced gene expression. Ha-ras, c-myc and p53 mRNA level were measured 48 hours after the DMBA induction in different organs (spleen, lung, thymus, kidney, liver and lymph node). The CBA/Ca mice received tap water (150 $\pm$ 3 ppm deuterium) in the control group and deuterium depleted water (21 $\pm$ 3 ppm deuterium) in the treated group for 48 hours.

were detected on X-ray film, developed, scanned into the computer and analysed by Quantiscan software (Biosoft, Cambridge, UK). The level of c-myc, Ha-ras and p53 gene expression was determined by percentage of positive control.

# **Results and Discussion**

It is known that, after carcinogen exposure of sensitive mice, an increase of certain (proto)oncogene and tumour suppressor gene expressions can be observed. The present study was meant to answer the question whether deuterium depletion can influence the early response of these genes, which have a central role in tumorigenesis. In order to cause deuterium depletion in mice, the drinking water (150 ppm  $\pm$  3 ppm) of the treated animals was replaced with deuterium depleted water (21 ppm  $\pm$  3 ppm ). The effect of deuterium depletion on c-myc, Ha-ras and p53 gene expression was studied after inducing the expression of these genes with DMBA. The rate of gene expression of these genes was determined in six different organs (spleen, lung, thymus, kidney, liver and lymph node) of the mice. The results summarized in Figure 1, show that independently of the organs or the genes, the expression was lower in the DDW-treated than in the control group.

The RNA expression pattern in the investigated organs was different. Further investigations are required to reveal a possible tissue specificity. The results of the present study, however, clearly indicate the inhibitory effect of deuterium depletion on the expression of these genes.

The molecular pathomechanism of carcinogenesis involves the changes of expression of oncogenes and tumor suppressor genes (31). Decreasing of (proto)oncogene and suppressor gene (over)expression may decrease the risk of malignant transformation (32-35). Since our earlier study (7-9) proved that deuterium depletion can result in tumour regression in mice, dogs, cats and in humans, we assume that the genes that have key role in tumorigenesis are sensitive to a shortage of deuterium.

Several studies have proved the effect of deuterium at high concentration which is the consequence of isotopic effect. Heavy water was found to accelerate the rate of actin polymerization and to increase microfilament gelation (36), to arrest cell proliferation (37) and also to inhibit heat shock protein induction (38).

We would like to emphasize that the application of DDW results in a decrease of deuterium concentration close to the natural level. The present results also seem to support the hypothesis that naturally occurring deuterlum may be involved in the regulation of genes which have a key role in cell cycle regulation and/or in tumour development.

### References

- 1 Collins CJA Bowman NS (eds.): Isotope Effects in Chemical Reactions. Van Nostrand Reinhold, New York, 1971.
- 2 Wiberg KB: The deuterium isotope effect. Chem Rev 55: 713-43, 1955.
- 3 Jancsó G and Van Hook WA: Condensed phase isotope effects

- (especially vapor pressure isotope effects). Chem Rev 74: 689-750, 1974.
- 4 Rundel PW, Ehleringer JR and Nagy KA: Stable isotopes in ecological research. Springer, New York, 1988.
- 5 Katz JJ and Crespi HL: Isotope effects in biological systems. *In:* Collins CJ, Bowman NS (eds.): Isotope effects in chemical reactions. Van Nostrand Reinhold, New York: 286-363, 1971.
- 6 Yurtsever Y and Gat JR: Stable isotope hydrology. Edited by Gat JR and Gonfiantini R: International Atomic Energy Agency, Vienna, 103-42, 1981.
- 7 Somlyai G, Jancsó G, Jákli G, Vass K, Barna B, Lakics V and Gaál T: Naturally occurring deuterium is essential for the normal growth rate of cells. FEBS Lett 317: 1-4, 1993.
- 8 Somlyai G, Láskay G, Berkényi T, Jákli and Jancsó G: Naturally occurring deuterium may have a central role in cell signalling. *In:* Heys JR, Melillo DG (eds.): Synthesis and applications of isotopically labelled compounds. John Wiley & Sons Ltd, 137-41, 1998.
- 9 Somlyai G, Laskay G, Berkényi T, Galbács Z, Galbács G, Kiss AS, Jákli Gy and Jancsó G: The biological effects of deuterium-depleted water, a possible new tool in cancer therapy. Z Onkol J Oncol 30: 91-94, 1998.
- 10 Ember I, Kiss I and Pusztai Zs: Effect of 7,12-dimethylbez(α)anthracene on onco/suppressor gene action in vivo: a short-term experiment. Anticancer Res 18: 445-7, 1998.
- 11 Pusztai Zs, Selypes A and Ember I: Short-term effects of 1-nitropyrene on chromosomes and on oncogene/tumor suppressor gene expression in vivo. Anticancer Res 18: 4489-92, 1998.
- 12 Ember I, Kiss I and Vermes E: Early effect of cyclophosphamide on oncogene expression in vivo. In Vivo 12: 201-8, 1998.
- 13 Ember I and Kiss I: *In vivo* effects of cyclophosphamide on oncogene and suppressor gene expression in a "follow-up study". Anticancer Res 17: 3593-7, 1997.
- 14 Ember I, Kiss I, Ghodratollah NA Raposa T: Effect of ABVD therapeutic protocol on oncogene and tumor suppressor gene expression in CBA/CA mice. Anticancer Res 18: 1149-52, 1998.
- 15 Ember I, Raposa T, Varga Cs, Herceg L and Kiss I: Carcinogenic effects of cytostatic protocols in CBA/Ca mice. In Vivo 9: 65-9, 1995.
- 16 Ember I, Raposa T, Varga Cs and Kiss I: Effect of different cytostatic protocols on oncogene expression in CBA/Ca mice. Anticancer Res 15: 1285-8, 1995.
- 17 Ember I, Kiss I, Raposa T, Nowrasteh G and Matolcsy A: In vivo effects of CHOP protocol on onco and suppressor gene expression: follow-up study. In Vivo 12: 489-94, 1998.
- 18 Ember I, Kiss I and Raposa T: In vivo effects of COPP protocol on onco- and suppressor gene expression in "follow-up study". In Vivo 11: 399-402, 1997.
- 19 Ember I, Kiss I and Malovics I: Oncogene and tumor suppressor gene expression changes in person exposed to ethylene oxide. Eur J Cancer Prev 7: 167-8, 1998.
- 20 Ember I, Kiss I, Gömbkötó Gy, Muller E and Szeremi M: Oncogene and suppressor gene expression as a biomarker for ethylene oxide exposure. Cancer Det Prev 22: 241-5, 1998.
- 21 Ember I, Kiss IA and Faluhelyi Zs: Gene expression changes as potential biomarkers of tumour bearing status in humans. Eur J Cancer Prev 7: 347-8, 1998.
- 22 Pusztai Zs, Dezsenyi T, Kiss T, Gecsei Gy and Ember I: Detection of

- elevated oncogene expressions in brain tumours and their macroscopically healthy surrounding tissues. Eur J Cancer Prev 7: 417-9, 1998.
- 23 Selypes A and Ember I: Investigation of the total RNA expression of the genes c-myc and p53 in the testes of mice treated with cadmium. Central Eur J Occup Env Med 4: 36-44, 1998.
- 24 Kozma L, Kiss I, Nagy A, Szakal S and Ember I: Investigation of c-myc and K-ras amplification in renal clear cell adenocarcinoma. Cancer Lett 111: 127-31, 1997.
- 25 Hajdu J, Kozma L, Kiss I, Szentkereszty Z, Szakall S and Ember I: Is the presence of distant metastasis associated with c-myc amplification in gastric cancer? Acta Chir Hung 36: 119-21, 1997.
- 26 Kozma L, Kiss I, Ember I and Kertai P: Studies on accute myelomonotic leukemia in LBFl rats. Cancer Lett 68: 185-92, 1993.
- 27 Kozma L, Kiss I, Szakall S and Ember I: Investigation of c-myc oncogene amplification in colorectal cancer. Cancer Lett 81: 165-9, 1994
- 28 Ember I, Kiss I and Nowrasteh G: Different H2 haplotypes have a strong influence on oncogene action. Anticancer Res 19: 1181-5, 1999.
- 29 Chomczynski P and Sacchi N: Single step method for RNA isolation by acidguanidinium thiocyanate-phenol-chloroform extraction. Anal Biochem 162: 156-9, 1987.
- 30 Renz M and Kurz C. A colorimetric method for DNA hybridization. Nucl Acids Res 12: 3435-44, 1984.
- 31 Brandt-Rauf PW: Biomarkers of gene expression: growth factors and oncoproteins. Environ Health Perspect *105*: 807-16, 1997.
- 32 MacArthur LH, Clarke MF and Westin EH: Malignant transformation of NIH 3T3 fibroblast by human c-sis is dependent upon the level of oncogene expression. Mol Carcinog 5: 311-9, 1992.
- 33 Rimoldi D, Srikantan V, Wilson VL, Bassin RH and Samid D: Increased sensitivity of nontumorigenic fibroblasts expressing ras or myc oncogenes to malignant transformation induced by 5-aza-2'deoxycytidine. Cancer Res 51: 324-30, 1991.
- 34 Robinson LA, Smith LJ, Fontaine MP, Kay HD, Mountjoy CP and Pirruccello SJ: c-myc antisense oligodeoxyribonucleotides inhibit proliferation of non-small cell lung cancer. Ann Thorac Surg 60: 1583-91, 1995.
- 35 Wang DG, Johnston CF and Buchanan KD: Oncogene expression in gastroenteropancreatic neuroendocrine tumors: implications for pathogenesis. Cancer 80: 668-75, 1997.
- 36 Omori H, Kuroda M, Naora H, Takeda H, Nio Y, Otani H and Tamura K: Deuterium oxide (heavy water) accelerates actin assembly *in vitro* and changes microfilament distribution in cultured cells. Eur J Cell Biol 74: 273-80, 1997.
- 37 Lamprecht J, Schroeter D and Pawaletz N: Mitosis arrested by deuterium oxide. Light microscopic, immunofluorescence and ultrastructural characterization. Eur J Cell Biol *51*: 303-12, 1990.
- 38 Edington BV, Whelan SA and Hightower LE: Inhibition of heat shock (stress) protein induction by deuterium oxide and glycerol: additional support for the abnormal protein hypothesis of induction J Cell Physiol 139: 219-28, 1989.

Received December 3, 1999 Accepted February 23, 2000